

1 **Role of the nucleotide excision repair pathway proteins (UvrB and UvrD2) in recycling**  
2 **UdgB, a base excision repair enzyme in *Mycobacterium smegmatis***

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4 *Running title: UvrB and UvrD2 facilitate turnover of AP-site bound UdgB*

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18 mycobacteria

## 19 S1. Supplementary Methods

20 **S1.1. Generation of constructs:** pJAM2 and its derivatives, pJAM\_FLAG and pJAMH\_His,  
21 were used to overexpress and purify proteins. To derive pJAM\_FLAG, a DNA sequence  
22 encoding FLAG-tag followed by a stop codon was introduced at XbaI restriction site, using  
23 oligomers UVL1 and UVL2. The plasmid pJAM\_FLAG retains a single XbaI restriction site  
24 before the FLAG-tag and His<sub>6</sub>-tag present after FLAG-tag is not translated. A hygromycin  
25 resistant version of pJAM2 plasmid, pJAMH\_His, was made by deleting Kan<sup>R</sup> cassette using  
26 NdeI restriction site. The vector fragment was end-filled by Klenow DNA polymerase, purified  
27 by phenol-chloroform extraction, and ligated with Hyg<sup>R</sup> cassette excised from pMV261<sup>hyg</sup> using  
28 HpaI and DraI restriction sites.

29 The PCR carried out in 20 µl volume contained 250 ng genomic DNA of *Msm* mc<sup>2</sup>155  
30 strain, 0.5 units of Q5 DNA polymerase (NEB), 1X Q5 reaction buffer, 1X GC enhancer, 250  
31 µM dNTPs, and 10 pmol each of the forward and reverse primers. Reaction conditions included  
32 an initial denaturation at 98°C for 2 min, followed by 30 cycles of 98 °C for 40 s, 50 – 60 °C for  
33 35 s and 72 °C for 1 - 3 min and a final extension at 72 °C for 5 min.

34 For cloning in one or more plasmids, pJAM2, pJAM\_FLAG and pJAMH\_His, ORFs  
35 (*MSMEG\_5031* (*Msm\_udgB*), *MSMEG\_5423* (*Msm\_mfd*), *MSMEG\_5534* (*Msm\_uvrD1*),  
36 *MSMEG\_1952* (*Msm\_uvrD2*), and *MSMEG\_2758* (*Msm\_mysA*)) from *Msm* mc<sup>2</sup>155 were PCR  
37 amplified using the respective primers with flanking BamHI or XbaI cloning sites (Table S1).  
38 Purified PCR products were ligated with the vector DNAs using BamHI and/or XbaI restriction  
39 sites, to obtain C-terminally His<sub>6</sub>-tagged or FLAG-tagged proteins. In parallel, pJAM2 harboring  
40 a C-terminally truncated variant of *Msm\_mfd*, encoding the first 1040 amino acids was also  
41 generated using *Msm\_mfd\_BamHI\_Fp* and *Msm\_mfd\_1040\_XbaI\_Rp* primers.

42 To generate pMV261h\_udgB, *Msm\_udgB* encoding ORF, *MSMEG\_5031*, was PCR  
 43 amplified using the phosphorylated primers *Msm\_udgB*\_PFp (binds 187 bp upstream of ORF)  
 44 and *Msm\_udgB*\_Rp (binds 123 bp downstream of ORF). PCR amplicon (1200 bp) was ligated  
 45 into PvuII digested pMV261hyg vector.

46 **Table S1.** List of DNA oligomers used in the study to generate constructs and verify strains

<b>DNA oligomer</b>	<b>Sequence (5' to 3'), restriction site is underlined</b>
<i>Msm_udgB</i> _PFp	GACACCGGTACCGGCGATCAGTG
<i>Msm_udgB</i> _Rp	CAGAAGCTTTGGTATCCGAGCCG
<i>Msm_udgB</i> _BamHI_Fp	CTGGTGGGATCCATGGGACCGATTTTC
<i>Msm_udgB</i> _XbaI_Rp	CGGTCATCTAGATTGCCCGTCACG
<i>Msm_uvrD1</i> _BamHI_Fp	TACGGATCCATGACTTCCCCC
<i>Msm_uvrD1</i> _XbaI_Rp	GGAGTCTAGAGAGCTTCTGCAG
<i>Msm_uvrD2</i> _BamHI_Fp	CGGTGTGGATCCTTGTCGGGCGGT
<i>Msm_uvrD2</i> _XbaI_Rp	GCAGCATCTAGACGAATCGTGGCGGTT
<i>Msm_mfd</i> _BamHI_Fp	ATCCAGGATCCATGACCGCACCG
<i>Msm_mfd</i> _XbaI_Rp	GCACCTTCTAGATCGCTTCGCCTC
<i>Msm_mfd</i> _1040_XbaI_Rp	GGACTTCTAGACGGTGTGGCAAC
<i>Msm_mysA</i> _XbaI_Fp	AGGCTCTAGAGTGGCAGCGACAAAG
<i>Msm_mysA</i> _XbaI_Rp	CATTTCTAGACTAGTCCAGGTAGTCGCG
UVL1	CTAGAGACTACAAGGACGACGACGACAAGTGAG
UVL2	CTAGCTCACTTGTCGTCGTCGTCCTTGTAGTCT
<i>MsuvrB</i> -Fp	CGCACCGGCAAACCCTTCG (binds 117 bps inside the ORF)
<i>MsuvrB</i> -Rp	CGCGAATTCAGTCACGACGAC (binds 50 bps downstream the ORF)

48 **S1.2. Generation of *Msm uvrB*<sup>-</sup> *udgB*<sup>-</sup> strain:** The plasmid, pPR-*Ms\_uvrB-kan*<sup>R</sup>, was  
49 electroporated in *Msm udgB*<sup>-</sup> for allelic exchange and the transformants were selected on LBT  
50 agar in the presence of Kan, Hyg and Gm at 30 °C. The transformants were inoculated in 2 ml of  
51 LBT Kan Hyg and incubated at 30 °C for 2 days. Bacterial cultures were serially diluted till 10<sup>-2</sup>,  
52 and aliquots of 50 µl from each serial dilution were plated on LBT agar supplemented with Kan,  
53 Hyg and 10% sucrose. Following incubation at 39 °C for 3 days, isolated colonies obtained on  
54 the plate were patched on LBT agar containing either Kan, Hyg and 10% sucrose or Hyg and  
55 Gm. Colonies that grew only on LBT agar plates containing Kan, Hyg and 10% sucrose were  
56 further verified by PCR using ORF primers, *MsuvrB*-Fp and *MsuvrB*-Rp.

### 57 **S1.3. Overexpression and purification of proteins**

58 **S1.3.1. *MsmUvrA*, *MsmUvrB* and *MsmUvrC*:** N-terminally His<sub>6</sub>-tagged NER proteins were  
59 overexpressed and purified from *Msm* [1].

60 **S1.3.2. *MsmUdgB*\_FLAG and *MsmUdgB*\_His<sub>6</sub>:** C-terminally FLAG-tagged UdgB (used for *in*  
61 *vitro* pulldown assays) was purified from *Msm* mc<sup>2</sup>155 strain harboring the pJAM\_*udgB*\_FLAG  
62 plasmid. A single colony from the transformation plate was inoculated in 2 ml LBT Kan and  
63 grown at 37 °C, 180 rpm for 2 days. LBT Kan (50 ml) was inoculated with 1% of saturated  
64 culture and incubated at 37 °C, 180 rpm for 12-14 h. Inoculation of 3 l LBT Kan was done with  
65 1% primary culture and incubated at 37 °C, 160 rpm till OD<sub>600</sub> reached 0.6. Protein was  
66 expressed using 0.5% acetamide for 4 h. The cell pellet was washed and resuspended in ice-cold  
67 25 ml lysis buffer (50 mM Na<sub>3</sub>PO<sub>4</sub> pH 7, 50 mM NaCl, 10% glycerol, and 2 mM β-  
68 mercaptoethanol). The following steps were performed at 4 °C. The cells were lysed by  
69 sonication and the lysate was subjected to centrifugation at 24K rpm, 4 °C for 1 h. The soluble  
70 cell-free extract (CFE) was loaded on heparin column (GE Healthcare), washed with 5 column

71 volumes of lysis buffer containing 200 mM NaCl and the protein was eluted with a gradient (0%  
72 to 100%) of NaCl in elution buffer [50 mM Na<sub>3</sub>PO<sub>4</sub> pH 7, 1 M NaCl, 10 % glycerol (v/v) and 2  
73 mM β-mercaptoethanol) at a flow rate of 1 ml/min for 30 min. The fractions eluted with 500 mM  
74 to 800 mM NaCl were enriched for the desired protein. The protein containing fractions were  
75 pooled and dialyzed to gradually obtain 50 mM NaCl concentration. The dialyzed fraction was  
76 loaded on Mono-S (GE Healthcare), washed with 3 column volumes of lysis buffer and the  
77 protein was eluted with a gradient of NaCl as explained above. Near homogenous protein  
78 fractions were pooled, dialyzed against buffer (50 mM Na<sub>3</sub>PO<sub>4</sub> pH 7, 150 mM NaCl, 50%  
79 glycerol, 2 mM β-mercaptoethanol), quantified and stored at -20 °C.

80 Similarly, C-terminally His<sub>6</sub>-tagged UdgB (used for *in vitro* enzymatic reactions) was purified  
81 from *Msm ung*- strain harboring the pJAMH\_udgB\_His plasmid. Protein was overexpressed as  
82 described above and purified as discussed before [2].

83 **S1.3.3. Translocases:** *MsmMfd*, *MsmMfd*\_1040 and *MsmUvrD2* were purified using the  
84 construct pJAM\_*mfd* or pJAM\_*mfd*\_1040 or pJAM\_*uvrD2*\_His, respectively in *Msm mc*<sup>2155</sup>.  
85 Proteins were expressed using 0.5% acetamide followed by 6-8 h incubation at 37 °C and 180  
86 rpm. Like UvrA, all three proteins were expressed in the soluble fraction and C-terminally His<sub>6</sub>-  
87 tagged proteins were purified by affinity chromatography (Ni-NTA) followed by size-exclusion  
88 chromatography.

89 **S1.3.4. *Msm*RNAP:** RNAP was purified from *Msm mc*<sup>2155</sup>/ pJAM\_*mysA* strain by  
90 overexpressing the sigma factor, SigA (σ<sup>A</sup>), to enrich the RNAP holoenzyme with σ<sup>A</sup> factor [3].

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92 **Table S2.** List of DNA oligomers used for biochemical assays

<b>DNA Oligomer</b>	<b>Sequence (5' to 3'), complementary sequence is underlined</b>	<b>Description</b>
GU9	<u>CTCAAGTG</u> <b>U</b> <u>AGGCATGCTTTTGC</u> <u>ATGCCTGCACTTGA</u>	37 nt long, makes a stem (16 bp) loop (4 nt) structure and contains G:U at 9 <sup>th</sup> position in stem
GU26	<u>GACTACGTA</u> <u>CTGTCACGCTCAAG</u> <u>TG</u> <b>U</b> <u>AGGCATGCATCAGGCCAGA</u> <u>TCTGCTTTT</u> <u>AGCAGATCTGGCC</u> <u>TGATGCATGCCT</u> <b>G</b> <u>CACTTGAGCG</u> <u>TGACAGTACGTAGTC</u>	106 nt long, makes a stem (51 bp) loop (4 nt) structure and contains G:U at 26 <sup>th</sup> position in stem
UVL3	CTACTACGTA <u>CTGTCAGGGGTCC</u> AT <b>U</b> TTCA <u>CCGGAATCAGGCCAG</u> ATCTGCTAGTCTAGAGGATGCTA AGGTC	73 nt long ssDNA containing uracil at 25 <sup>th</sup> position
UVL4	GCAGATCTGGCCTGATTCCGGTG AAGATGGACCCCTGACAGTACG TAGTAG	51 nt long and complementary to UVL3
5' Biotinylated SSU9	C( <b>B</b> )TCAAGTG <b>U</b> AGGCATGCAAG AGCT	24 nt long ssDNA harboring uracil at 9 <sup>th</sup> position and biotin modification at 5' end
UVL9	AGCTCTTGCA <u>TGCCTGCACTTGA</u> GTAGTCTAGAGGATGC	39 nt long and complementary to biotinylated SSU9
5' Biotinylated oligomer	GAC( <b>B</b> )GCTGCCGAATTCTGGCTT GCTAAAGGATAGTCGAATTTTCT CATTTT	51 nt long ssDNA with biotin modification at 5' end [4]

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## S2. Supplementary Results

### S2.1. Raw data for growth curves

NOTE: Values starred (\*) and highlighted in blue color were excluded while plotting the growth curves.

Time (hours)	<i>Msm</i> WT /pMV261h (U)				<i>Msm</i> WT /pMV261h (2.5 mM H <sub>2</sub> O <sub>2</sub> )			
0	0.106	0.103	0.102	0.105	0.11	0.114	0.107	0.108
3	0.148	0.157	0.156	0.161	0.166	0.176	0.156	0.16
6	0.148	0.153	0.147	0.152	0.135	0.143	0.132	0.132
9	0.197	0.201	0.199	0.206	0.152	0.169	0.161	0.161
12	1.533*	0.302	0.287	0.29	0.196	0.224	0.199	0.203
15	0.432	0.474	0.456	0.458	0.299	0.347	0.317	0.315
18	0.645	0.717	0.691	0.686	0.463	0.542	0.485	0.495
21	0.863	0.915	0.896	0.89	0.665	0.778	0.766	0.759
24	1.047	1.099	1.083	1.083	0.882	0.979	0.964	0.956
27	1.184	1.237	1.216	1.232	1.078	1.163	1.144	1.157
30	1.263	1.302	1.287	1.304	1.241	1.28	1.268	1.293
33	1.312	1.34	1.313	1.323	1.318	1.336	1.342	1.363
36	1.317	1.35	1.321	1.346	1.349	1.353	1.36	1.379
39	1.327	1.359	1.317	1.342	1.369	1.374	1.372	1.397
42	1.32	1.339	1.318	1.333	1.377	1.372	1.382	1.411
45	1.336	1.345	1.317	1.33	1.359	1.361	1.369	1.387
48	1.321	1.339	1.312	1.328	1.363	1.364	1.364	1.38

Time (hours)	<i>Msm</i> WT /pMV261h_udgB (U)				<i>Msm</i> WT /pMV261h_udgB (2.5 mM H <sub>2</sub> O <sub>2</sub> )			
0	0.106	0.101	0.105	0.104	0.112	0.103*	0.11	0.114
3	0.162	0.153	0.167	0.162	0.166	0.143*	0.157	0.162
6	0.149	0.126	0.139	0.137	0.138	0.095*	0.121	0.125
9	0.195	0.154	0.174	0.171	0.168	0.094*	0.138	0.143
12	0.243	0.19	0.208	0.209	0.207	0.088*	0.162	0.165
15	0.335	0.272	0.27	0.283	0.281	0.094*	0.202	0.214
18	0.484	0.415	0.37	0.409	0.393	0.086*	0.254	0.273
21	0.691	0.648	0.522	0.58	0.584	0.092*	0.341	0.406
24	0.884	0.864	0.768	0.823	0.798	0.091*	0.495	0.612
27	1.012	1.025	0.939	0.979	1.001	0.090*	0.787	0.861
30	1.104	1.139	1.053	1.098	1.152	0.086*	0.964	1.031
33	1.166	1.197	1.159	1.188	1.25	0.094*	1.125	1.169
36	1.191	1.22	1.193	1.225	1.292	0.088*	1.235	1.256
39	1.206	1.228	1.211	1.238	1.317	0.089*	1.297	1.315
42	1.201	1.229	1.207	1.246	1.329	0.092*	1.33	1.33
45	1.201	1.226	1.217	1.234	1.327	0.083*	1.326	1.332
48	1.211	1.226	1.223	1.237	1.34	0.088*	1.333	1.347

Time (hours)	<i>Msm uvrB</i> <sup>-</sup> /pMV261h (U)				<i>Msm uvrB</i> <sup>-</sup> /pMV261h (2.5 mM H <sub>2</sub> O <sub>2</sub> )			
0	0.106	0.109	0.111	0.111	0.112	0.11	0.11	0.115
3	0.162	0.156	0.169	0.175	0.165	0.169	0.169	0.171
6	0.14	0.143	0.149	0.154	0.133	0.137	0.129	0.143
9	0.184	0.18	0.185	0.185	0.158	0.163	0.146	0.168

12	0.245	0.234	0.256	0.245	0.19	0.195	0.169	0.195
15	0.38	0.351	0.404	0.382	0.258	0.262	0.222	0.279
18	0.581	0.528	0.612	0.573	0.359	0.361	0.292	0.363
21	0.751	0.712	0.769	0.735	0.546	0.54	0.427	0.555
24	0.894	0.833	0.909	0.844	0.732	0.741	0.625	0.703
27	1	0.969	1.022	0.963	0.849	0.9	0.766	0.834
30	1.089	1.067	1.09	1.067	0.998	1.047	0.91	0.98
33	1.13	1.112	1.137	1.122	1.125	1.149	1.054	1.09
36	1.152	1.142	1.159	1.143	1.167	1.181	1.115	1.125
39	1.149	1.141	1.148	1.127	1.199	1.219	1.149	1.141
42	1.14	1.135	1.136	1.127	1.206	1.224	1.159	1.159
45	1.123	1.121	1.112	1.089	1.196	1.217	1.154	1.149
48	1.106	1.123	1.107	1.068	1.19	1.2	1.16	1.157

Time (hours)	<i>Msm uvrB</i> <sup>-</sup> /pMV261h_ <i>udgB</i> (U)				<i>Msm uvrB</i> <sup>-</sup> /pMV261h_ <i>udgB</i> (2.5 mM H <sub>2</sub> O <sub>2</sub> )			
0	0.109	0.107	0.109	0.105	0.107	0.109	0.109	0.106
3	0.158	0.168	0.179	0.167	0.158	0.156	0.16	0.159
6	0.143	0.144	0.153	0.136	0.11	0.122	0.135	0.11
9	0.151	0.154	0.168	0.147	0.116	0.13	0.149	0.830*
12	0.177	0.178	0.198	0.175	0.117	0.134	0.176	0.121
15	0.234	0.236	0.267	0.229	0.141	0.166	0.205	0.133
18	0.331	0.327	0.36	0.316	0.141	0.167	0.239	0.132
21	0.472	0.464	0.481	0.463	0.169	0.208	0.306	0.147
24	0.633	0.644	0.647	0.653	0.176	0.273	0.457	0.167
27	0.749	0.754	0.716	0.723	0.218	0.386	0.619	0.161
30	0.847	0.859	0.834	0.816	0.284	0.544	0.689	0.188
33	0.961	0.988	0.945	0.901	0.451	0.681	0.803	0.263
36	1.055	1.062	1.033	1.005	0.602	0.768	0.898	0.341
39	1.058	1.067	1.05	1.011	0.67	0.851	0.967	0.487
42	1.064	1.078	1.055	1.026	0.756	0.955	0.994	0.588
45	1.068	1.058	1.045	0.997	0.794	0.983	1.011	0.651
48	1.064	1.074	1.039	1.003	0.861	0.982	1.003	0.735

Time (hours)	<i>Msm uvrB</i> <sup>-</sup> <i>udgB</i> <sup>-</sup>				<i>Msm uvrB</i> <sup>-</sup> <i>udgB</i> <sup>-</sup> (2.5 mM H <sub>2</sub> O <sub>2</sub> )			
0	0.114	0.114	0.118	0.118	0.113	0.115	0.115	0.114
3	0.19	0.192	0.193	0.196	0.17	0.187	0.181	0.181
6	0.182	0.182	0.181	0.182	0.163	0.167	0.162	0.151
9	0.262	0.249	0.249	0.264	0.218	0.22	0.216	0.212
12	0.435	0.368	0.365	0.429	0.301	0.297	0.3	0.303
15	0.737	0.636	0.597	0.731	0.505	0.484	0.476	0.512
18	0.927	0.874	0.848	0.936	0.789	0.741	0.737	0.801
21	1.135	1.06	1.063	1.152	0.986	0.949	0.894	1.006
24	1.274	1.232	1.216	1.282	1.151	1.143	0.929	1.199
27	1.336	1.302	1.291	1.336	1.268	1.277	0.87	1.325
30	1.355	1.316	1.307	1.353	1.31	1.324	0.862	1.368
33	1.347	1.32	1.308	1.346	1.34	1.36	0.865	1.395
36	1.345	1.324	1.328	1.345	1.345	1.369	0.861	1.404
39	1.296	1.281	1.269	1.304	1.338	1.365	0.867	1.391
42	1.29	1.271	1.26	1.286	1.353	1.359	0.866	1.388
45	1.023	1.191	1.212	1.158	1.32	1.33	0.846	1.297



48	0.877	1.098	1.192	1.018	1.315	1.314	0.847	1.06
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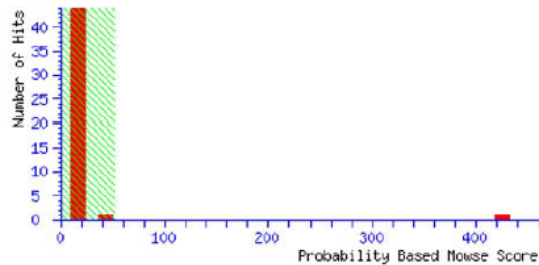
<b>Time (hours)</b>	<b>Media Control</b>			
0	0.064	0.064	0.064	0.064
3	0.063	0.063	0.063	0.063
6	0.064	0.064	0.064	0.064
9	0.066	0.066	0.066	0.066
12	0.066	0.066	0.066	0.066
15	0.068	0.068	0.068	0.068
18	0.068	0.068	0.068	0.068
21	0.068	0.068	0.068	0.068
24	0.068	0.068	0.068	0.068
27	0.068	0.068	0.068	0.068
30	0.068	0.068	0.068	0.068
33	0.068	0.068	0.068	0.068
36	0.068	0.068	0.068	0.068
39	0.067	0.067	0.067	0.067
42	0.068	0.068	0.068	0.068
45	0.068	0.068	0.068	0.068
48	0.068	0.068	0.068	0.068

## S2.2. Raw MS analysis of interacting proteins (trypsin digested and subjected to MALDI) identified in pulldown assays

### Protein band 1, Table 3

#### Probability Based Mowse Score

Ions score is  $-10 \cdot \log(P)$ , where P is the probability that the observed match is a random event. Protein scores greater than 51 are significant ( $p < 0.05$ ).



#### Concise Protein Summary Report

[Switch to full Protein Summary Report](#)

To create a bookmark for this report, right click this link: [Concise Summary Report \(../data/20210609/F024413.dat\)](#).

Re-Search All

Search Unmatched

1. [MSMEG\\_1368](#) Mass: 147106 Score: 425 Expect: 2.1e-039 Peptides matched: 56  
DNA-directed RNA polymerase, beta' subunit (rpoC) [2.7.7.6] {Mycobacterium smegmatis MC2 155}

Sequence Coverage: 53%

Matched peptides shown in **Bold Red**

```
1 MLDVNFDEL RIGLATADDI RNWSYGEVKK PETINRYTLK PEKGLFCEK
51 IFGPTRDWEK YCGKYKRVRF KGIIICERCGV EVTRAKVRRK RMGHIELAAP
101 VTHIWFYKGV PSRLGYLLDL APKDLEKIIY FAAYVITSVD DEMRHNELST
151 LEAEMAVEKK AVEDQRDADL EARAQKLEAD LAELEAEGAK SDVRRKVRDS
201 GEREMRQLRD RAQRELDRLD EIWNFTFKLA PKQLIVDEVL YRELQDRYGE
251 YFTGAMGAES IKKLIENFDI DAAEASLREV IRSKGQKQKL RALKRLKQVA
301 AFQQSGNSPM GMVLDAVPII PPELRPMVQL DGGRAFATSDL NDLYRRVINR
351 NNRLKRLIDL GAPEIIVNNE KRMLQESVDA LFDNGRGRPR VTGPGRNRLK
401 SLSDLLKGGK GRFRQNLGK RVDYSGRSVI VVGPQLKLHQ CGLPKLMALE
451 LFKPFVMKRL VDLNHAQNIK SAKRMVERQR PQVQVLEEV IAEHPVLLNR
501 APTLHRLGIQ AFEPQLVEGK AIQLHPLVCE AFNADFQGDQ MAVHLPLSAE
551 AQAEARILML SSNNILSPAS GKPLAMPRLD MVTGLYYLTT LVEGATGEYQ
601 AATKDAPEQG VYSSPAEAIM AMDRGALSVR AKIKVRLTEL RPPDTLEAQL
651 FENGWPGDA WTAETTLGRV MFNELLPKSY PFVNEQMHKK VQARIINDLA
701 ERFPMIVVAQ TVDKLKDAGF YWATRSQVTV SMADVLVPPQ KQEILERHEA
751 EADAIERKYQ RGALNHTERN ESLVKIWQDA TEEVGKALEE FYPADNPIIT
801 IVKSGATGNL TQTRTLAGMK GLVTNPKGEF IPRPKSSFR EGLTVLEYFI
851 NTHGARKGLA DTALRTADSG YLTRLVDVVS QDVIVREHDC ETERGINVTL
901 AERGPDGLI RDAHVETSFAF ARTLATDAVD ANGWIIERG HDLGDPAIDA
951 LLAAGITTVK VRSVLTCTSA TGVCAMCYGR SMATGKLVDI GEAVGI VAAQ
1001 SIGEPGQTL MRTFHQGGVT GGADIVGGLP RVQELFEARV PRNKAPIADV
1051 AGRVRL EESD KFFKITIVPD DGGEEVYVDK LSKRQRLRVI THEDGTEGVL
1101 SDGDHMEVGD QLMGAADPH EVLRVQGPRE VQIHLVKVQ EVYRAQGVSI
1151 HDKHIEVIVR QMLRVRTIID SGSTEFPLGS LTERAEFEAE NRRVVAEGGE
1201 PAAGRPLVMG ITKASLATDS WLSAASFQET TRVLTDAAIN CRSKLNGLK
1251 ENVIIGKLIIP AGTGISRYRN IQVQPTTEAR AAAYTIPSYE DQYSPDFGQ
1301 ATGAAVPLDD YGYSYDR
```

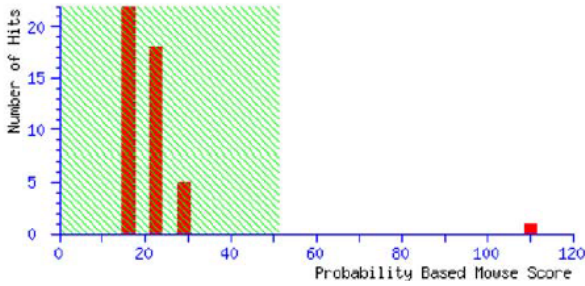
#### Search Parameters

Type of search : Peptide Mass Fingerprint  
Enzyme : Trypsin  
Fixed modifications : Carbamidomethyl (C)  
Variable modifications : Oxidation (M)  
Mass values : Monoisotopic  
Protein Mass : Unrestricted  
Peptide Mass Tolerance :  $\pm 0.5$  Da  
Peptide Charge State : 1+  
Max Missed Cleavages : 0

# Protein band 2, Table 3

## Probability Based Mowse Score

Ions score is  $-10 \cdot \log(P)$ , where P is the probability that the observed match is a random event. Protein scores greater than 51 are significant ( $p < 0.05$ ).



## Concise Protein Summary Report

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To create a bookmark for this report, right click this link: [Concise Summary Report \(../data/20210609/F024415.dat\)](#)

- [MSMEG\\_6091](#)      Mass: 93496      Score: **110**      Expect: 6.7e-008      Peptides matched: 26  
 negative regulator of genetic competence ClpC-mecB {Mycobacterium smegmatis MC2 155}

Sequence Coverage: 36%

Matched peptides shown in **Bold Red**

```

1 MFERFTDRAR RVVVLAQEEA RMLNHNHYIGT EHILLGLIHE GEGVAAKSLE
51 SLGISLEGVR SQVEEIIQQG QQAPSGHIPF TPRAKKVLLEL SLREALQLGH
101 NYIGTEHILL GLIREGEGVA AQVLVKLGAE LTRVRQQVIQ LLSGYQGKEA
151 AEAGTGGRRG ESGNPSTSLV LDQFGRNLTA AAMEGKLDPV IGREKEIERV
201 MQVLSRRTKN NPVLIGEPGV GKTAVVEGLA QAIIVHGEVPE TLKDKQLYTL
251 DLGSLVAGSR YRGDFEERLK KVLKEINTRG DIILFIDEUH TLVGAGAAEG
301 AIDAASILKP KLRARGELQTI GATTLDEYRK YIEKDAALER RFQPVQVGEP
351 TVEHTIEILK GLRDRYEAAH RVSITDSAMV AAATLADRYI NDRFLPDKAI
401 DLIDEAGARM RIRRMTAPPD LREFDEKIAD ARREKESAIQ AQDFEKAAL
451 RDKEKQLVAQ RAEREKQWRS GDLDVVAEVD DEQIAEVLGN WTIIPVFKLT
501 EETTRLLRM EELHKRIIG QEDAVKAVSK AIRRTRAGLK DPKRPSGSFI
551 FAGPSGVGKT ELSKALANFL FGDDALIQT DMGEFHDRFT ASRLFGAPPG
601 YVGEEGGQL TEKVRRRKPFV VVLFDEIEKA HQEIYNSLLQ VLEDGRLTDTG
651 QGRVTDFKNT VLIFTSNLGT SDISKAVGLG FSQGGSENNY ERMKQKVHDE
701 LKKHFRPEFL NRIDDIIIVFH QLTQDEIIQM VDLMIGRVSN QLTKDMMALE
751 LSDKAKALLA KRGDPVLGA RPLRRTIQRE IEDQLSEKIL FEEIGPGQLV
801 TVDVEGWGGE GQGEDAKFTF SGGPKRAETA EPDLAGAGAA GAPTAGTE

```

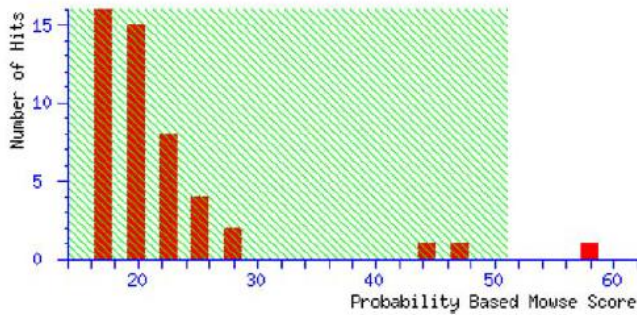
## Search Parameters

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 Enzyme : Trypsin  
 Fixed modifications : Carbamidomethyl (C)  
 Variable modifications : Oxidation (M)  
 Mass values : Monoisotopic  
 Protein Mass : Unrestricted  
 Peptide Mass Tolerance :  $\pm 0.5$  Da  
 Peptide Charge State : 1+  
 Max Missed Cleavages : 1

# Protein band 3, Table 3

## Probability Based Mowse Score

Ions score is  $-10 \cdot \log(P)$ , where P is the probability that the observed match is a random event. Protein scores greater than 51 are significant ( $p < 0.05$ ).



## Concise Protein Summary Report

[Switch to full Protein Summary Report](#)

To create a bookmark for this report, right click this link: [Concise Summary Report \(../data/20210609/F024435.dat\)](#)

1. [MSMEG\\_0005](#) Mass: 74580 Score: **58** Expect: 0.011 Peptides matched: 11  
DNA gyrase, B subunit (gyrB) [5.99.1.3] {Mycobacterium smegmatis MC2 155}

Sequence Coverage: 23%

Matched peptides shown in **Bold Red**

```
1 MAAQKNNAPK EYGADSITIL EGLEAVRKRP GMYIGSTGER GLHHLIWEVV
51 DNAVDEAMAG FATRVDKIIH ADGSVEVRDD GRGIPVEMHA TGMPTIDVVM
101 TQLHAGGKFD GETYAVSGGL HGVGSVWNA LSTRLEATVL RDGYEWFQYY
151 DRSVPGKLKQ GGETKETGTT IRFWADPEIF ETTDYNFETV ARRLQEMAFL
201 NKGLTIELTD ERVTAAEEVVD DWKDTAEAP KTADEKAAEA TGPSKVKKHRV
251 FHYPGGLVDY VKHINRTKTP IQQSIIDFDG KGPGHEVEIA MQWNAGYSES
301 WHTFANTINT HEGGTHEEGF RAALTSVWNR YAKDKLKLD KDPNLTGDDI
351 REGLAAVISV KVAEPQFEGQ TKTKLGNTEV KSEFVQKICNE QLQHWFEANP
401 AEAKTVVNKA VSSAQARIAA RKARELVRRK SATDIGGLPG KLADCRSTDP
451 SKSELYVVEG DSAGGSAKSG RDSMFQAILP LRGKIINWEK ARIDRVLKNT
501 EVQSIITALG TGIHDEFDIS KLRYHKIVLM ADADVDGQHI STLLLLLFR
551 FMKPLVENGH IFLAQPPLYK LKWQRSEPEF AYSDRERDGL LEAGRAAGKK
601 INVDDGIQRY KGLGEMDAKE LWETTMDPSV RVLRQVTLDD AAADELFSI
651 LMGEDVEARR SFITRNAKDV RFLDV
```

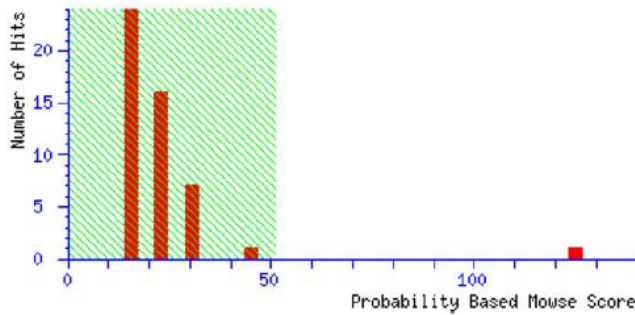
## Search Parameters

Type of search : Peptide Mass Fingerprint  
Enzyme : Trypsin  
Fixed modifications : Carbamidomethyl (C)  
Variable modifications : Oxidation (M)  
Mass values : Monoisotopic  
Protein Mass : Unrestricted  
Peptide Mass Tolerance :  $\pm 0.4$  Da  
Peptide Charge State : 1+  
Max Missed Cleavages : 0

# Protein band 4, Table 3

## Probability Based Mowse Score

Ions score is  $-10 \cdot \log(P)$ , where P is the probability that the observed match is a random event. Protein scores greater than 51 are significant ( $p < 0.05$ ).



## Concise Protein Summary Report

[Switch to full Protein Summary Report](#)

To create a bookmark for this report, right click this link: [Concise Summary Report \(.../data/20210609/F024419.dat\)](#)

- 1. [MSMEG\\_0880](#) Mass: 60472 Score: **125** Expect: 2.1e-009 Peptides matched: 15  
chaperonin GroL (groL) {Mycobacterium smegmatis MC2 155}

Sequence Coverage: 42%

Matched peptides shown in **Bold Red**

```
1 MRPGPHLREH TPSRPSRALH LTTTERAIPNP EEHFAMAKTI AYDEEARRGL
51 ERGLNSLADA VKVTLGPKGR NVVLEKKWGA PITINDGVSI AKEIELEDPY
101 EKIGAELVKE VAKKTDDVAG DGTTTATVLA QALVREGLRN VAAGANPLGL
151 KRGIEKAVEK VTETLLKSAK EVETKEQIAA TAGISAGDQS IGDLIAEAMD
201 KVGNEGVITV EESNTFGLQL ELTEGMRFDK GYISGFVTD AERQEAVLED
251 PYILLVSSKV STVKDLLLPLL EKVIQSGKPL LIAEDVEGE ALSTLVNKI
301 RGTFKSVAVK APGFGDRRRKA MLQDMAILTG QQVIESEEVGL SLETADVSLL
351 GKARKVVVTK DETTIVEGAG DAEAIQGRVA QIRAEIENSD SDYDREKLQE
401 RLAKLAGGVA VIKAGAATEV ELKERKHRIE DAVRNAKAAV EEGIVAGGV
451 ALLQSAPSLE ELSLTGDEAT GANIVRVALS APLKQIALNG GLEPGVVAEK
501 VSNLPAGHGL NAATGEYEDL LAAGVADPVK VTRSALQNAA SIAALFLTTE
551 AVVADKPEKA AAPAGDPTGG MGGMDF
```

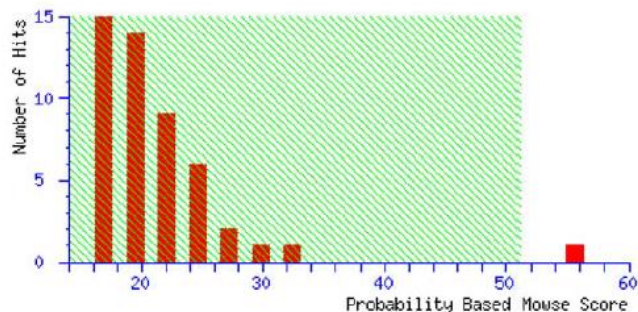
## Search Parameters

Type of search : Peptide Mass Fingerprint  
Enzyme : Trypsin  
Fixed modifications : Carbamidomethyl (C)  
Variable modifications : Oxidation (M)  
Mass values : Monoisotopic  
Protein Mass : Unrestricted  
Peptide Mass Tolerance :  $\pm 0.29$  Da  
Peptide Charge State : 1+  
Max Missed Cleavages : 0

## Protein band 5, Table 3

### Probability Based Mowse Score

Ions score is  $-10 \cdot \log(P)$ , where P is the probability that the observed match is a random event. Protein scores greater than 51 are significant ( $p < 0.05$ ).



### Concise Protein Summary Report

[Switch to full Protein Summary Report](#)

To create a bookmark for this report, right click this link: [Concise Summary Report \(./data/20210609/F024420.dat\)](#)

Re-Search All

Search Unmatched

1. [MSMEG\\_2430](#) Mass: 54788 Score: **56** Expect: 0.018 Peptides matched: 9  
signal recognition particle protein (ffh) {Mycobacterium smegmatis MC2 155}

Sequence Coverage: 20%

Matched peptides shown in **Bold Red**

```
1 MFESLSDRLT GALQGLRGKG RLTADADIDAT TREIRLALLE ADVSLPVRRA
51 FVARIKERAK GAEVSAALNP AQQVVKIVNE ELIGILGGET RQLAFAKTPP
101 TVIMLAGLQG AGKTTLAGKL AKWLKDKGHS PLLVACDLQR PGAVNQLQIV
151 GERAGVAVFA PHPGTAPGAH ETEGAGPGDP VAVAAAGLAE AKTKLYGVVI
201 VDTAGRLGID DVLMQAAGI RDAVQPDEL FVLDAMIGQD AVATAEAFRE
251 GVGFTGVVLT KLDGDARGGA ALSVREVTGV PILFASAGEK LEDFDVFHPD
301 RMASRILGMG DVLTLIEQAE QVFDQAQAE AAAKIGTGEL TLEDFLEQML
351 TIRKMGPIGN LLGMLPGAGQ MKDALAAVDD KQLDRVQAII RGMTPQERAD
401 PKIINASRRR RIANGSGVTV SEVNQLVDRF FEARKMMSM AGQMGMGFGR
451 KSATRKAAS KKKGKNAKK GKGPQPKVR NPLGAAGLPG GFPDLSNMPK
501 GLDELPPGLA DFDSLKLKFP NK
```

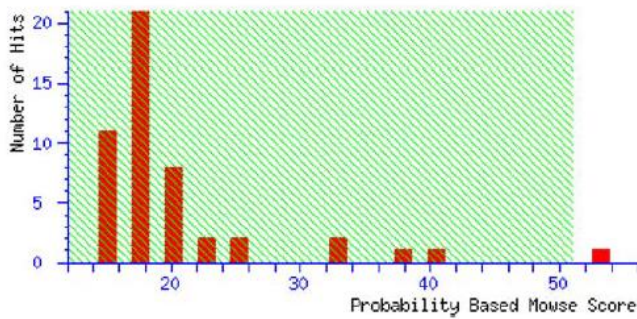
### Search Parameters

Type of search : Peptide Mass Fingerprint  
Enzyme : Trypsin  
Fixed modifications : Carbamidomethyl (C)  
Variable modifications : Oxidation (M)  
Mass values : Monoisotopic  
Protein Mass : Unrestricted  
Peptide Mass Tolerance :  $\pm 0.27$  Da  
Peptide Charge State : 1+  
Max Missed Cleavages : 0

## Protein band 6, Table 3

### Probability Based Mowse Score

Ions score is  $-10 \cdot \log(P)$ , where P is the probability that the observed match is a random event. Protein scores greater than 51 are significant ( $p < 0.05$ ).



### Concise Protein Summary Report

[Switch to full Protein Summary Report](#)

To create a bookmark for this report, right click this link: [Concise Summary Report\(../data/20210609/F024421.dat\)](#).

Re-Search All

Search Unmatched

1. [MSMEG\\_2389](#) Mass: 21217 Score: **53** Expect: 0.032 Peptides matched: 8  
DNA-binding protein HU (hup) {Mycobacterium smegmatis MC2 155}

Sequence Coverage: 42%

Matched peptides shown in **Bold Red**

```
1 MNKAEIDVL TTKMGDRRQ ATAAVENVVD TIVRAVHKGD SVTITGFGVF
51 EQRRRAARVA RNPRTGETVK VKPTSVPAFR PGAQFKAVIS GAQKLPADGP
101 AVKRGVTAGP AKKAAKKAPA KKAAAAKTAT KAAAKKAPAK KAATKAPAKK
151 AATKAPAKKA ATKAPAKKAA TKAPAKKAAA KAPAKKAATK APAKKAAAKK
201 APAKKGRR
```

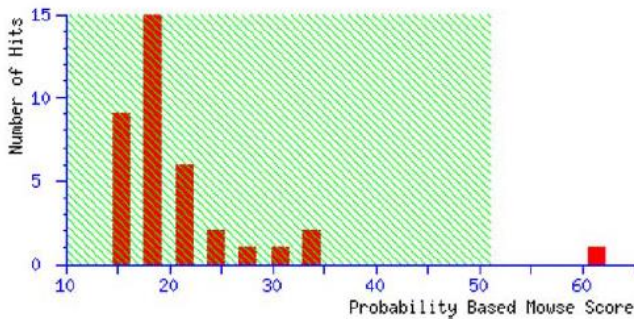
### Search Parameters

Type of search : Peptide Mass Fingerprint  
Enzyme : Trypsin  
Fixed modifications : Carbamidomethyl (C)  
Variable modifications : Oxidation (M)  
Mass values : Monoisotopic  
Protein Mass : Unrestricted  
Peptide Mass Tolerance :  $\pm 200$  ppm  
Peptide Charge State : 1+  
Max Missed Cleavages : 1

## Protein band 7, Table 3

### Probability Based Mowse Score

Ions score is  $-10 \cdot \log(P)$ , where P is the probability that the observed match is a random event. Protein scores greater than 51 are significant ( $p < 0.05$ ).



### Concise Protein Summary Report

[Switch to full Protein Summary Report](#)

To create a bookmark for this report, right click this link: [Concise Summary Report \(../data/20210609/F024412.dat\)](#)

Re-Search All

Search Unmatched

1. [MSMEG\\_6092](#) Mass: 12446 Score: **61** Expect: 0.005 Peptides matched: 6  
Lsr2 protein {Mycobacterium smegmatis MC2 155}

Sequence Coverage: 33%

Matched peptides shown in **Bold Red**

```
1 MAKKVTITLV DDFDGEATAD ETVEFGLDGV TYEIDLSAKN AAKLRNDLKQ
51 WVEAGRRVGG RKRGRAATTT TRGRGAIDRE QSAAIREWAR RRGHNVSTRG
101 RIPADVIDAF HAAT
```

### Search Parameters

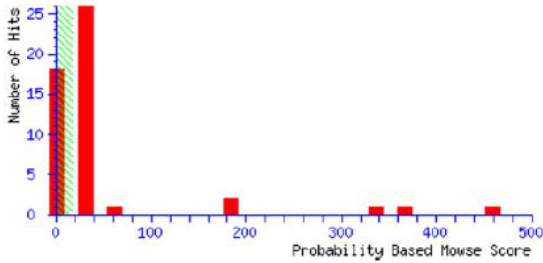
Type of search : Peptide Mass Fingerprint  
Enzyme : Trypsin  
Fixed modifications : Carbamidomethyl (C)  
Variable modifications : Oxidation (M)  
Mass values : Monoisotopic  
Protein Mass : Unrestricted  
Peptide Mass Tolerance :  $\pm 0.5$  Da  
Peptide Charge State : 1+  
Max Missed Cleavages : 1



# Protein band 1, Table 4

## Probability Based Mowse Score

Ions score is  $-10 \cdot \log(P)$ , where P is the probability that the observed match is a random event. Individual ions scores > 17 indicate identity or extensive homology ( $p < 0.05$ ). Protein scores are derived from ions scores as a non-probabilistic basis for ranking protein hits.



## Peptide Summary Report

[Switch to Protein Summary Report](#)

To create a bookmark for this report, right click this link: [Peptide Summary Report \(./data/20210608/F024380.dat\)](#)

Error tolerant

1. [MSMEG\\_3808](#) Mass: 109775 Score: 459 Peptides matched: 49  
excinuclease ABC, A subunit (uvrA) {Mycobacterium smegmatis MC2 155}

Sequence Coverage: 44%

Matched peptides shown in **Bold Red**

```
1  MGNVCVMRGT EHVRTACLDP RRPGRKPLAD RLVVKGAREH NLRGVLDLDP
51 RDALIVFTGL SGSGKSLAF DTIFAEQQR YVESLSAYAR QFLGQMDKPD
101 VDFIEGLSPA VSIDQKSTNR NPRSTVGTIT EVYDYLRLLY ARAGTPHCVP
151 CGERIARQTP QQIVDQVLAM DEGLRFQVLA PVVRTRKGEF VDLFEKLSNQ
201 GYSRVRVGV VYPLTDPPKL KKQEKHDIEV WDRLTVKAS AKQRLTDSIE
251 TALNLADGIV VLEFVDREDD HPHREQRFSE KLACPNHGHL AVDDLEPRSF
301 SFNSPYGACP ECTGLGIRKE VDPDLVVPDP ELTLAEGAVA PWSVGQSAEY
351 FTRMLAGLGE EMGFVNTPW KKLPAKARRA ILEGCDHQVH VRYKNRYGRT
401 RSYYADFEGV MAFLQRRMEQ TDSEQMKERY EGFMRDIPCP ECNGTRLKPE
451 ILAVTLASAGD FGAKSIAQVA ELSIADCADF LNSLTLGPRE QAIGAQVLKE
501 IQSRLGFLLD VGLDYLSLSR AAATLGGGEA QRIRLATQIG SGLVGVLYVL
551 DEPSIGLHQR DNRRLIDTLV RLRDLGNTLI WEHDLDTIA HADWVVDIGP
601 AAGEHGGQIV HSGTYDILLR NPESLTGAYL SGKESIEVPA IRRPVDDKRRQ
651 ITVWGAREN LKEIDVAFPL GVLTSVTGVS SGSKSTMVND ILATVLANKL
701 NGARLVPRGH TRVNGLDQLD KLVRVDQSPI GRTPRSNPAT YTGVPDKIRS
751 LFAATTEAKV RGYQPGRFSF NVKGGRCEAC SGDGTIKIEM NFLPDVYVPC
801 EVCHGARVNR ETLEVIHYGK TISEVLDMSI EEATEFFEPI SSIHRYLKTL
851 VDVGLGYVRL GQPAPTLSSG EAQRVKLAAE LQKRSTGRTI YILDEPTTGL
901 HFEDIRKLLK VINGLVDKGN TVIVIEHNLD VIKTSDWIID MGPEGGAGGG
951 TVVAQGTPEP VAANPDSYTG KFLAELLDVP TPKRKRKRKVS A
```

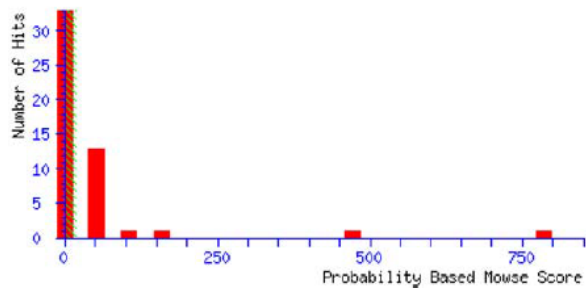
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Enzyme : Trypsin  
Fixed modifications : Carbamidomethyl (C)  
Variable modifications : Oxidation (M)  
Mass values : Monoisotopic  
Protein Mass : Unrestricted  
Peptide Mass Tolerance :  $\pm 0.01$  %  
Fragment Mass Tolerance :  $\pm 0.5$  Da  
Max Missed Cleavages : 0

# Protein band 1, Table 4

## Probability Based Mowse Score

Ions score is  $-10 \cdot \log(P)$ , where P is the probability that the observed match is a random event. Individual ions scores > 18 indicate identity or extensive homology ( $p < 0.05$ ). Protein scores are derived from ions scores as a non-probabilistic basis for ranking protein hits.



## Peptide Summary Report

[Switch to Protein Summary Report](#)

To create a bookmark for this report, right click this link: [Peptide Summary Report \(./data/20210608/F024376.dat\)](#)

**Error tolerant**

1. [MSMEG\\_1439](#) Mass: 30396 Score: **786** Peptides matched: 64  
ribosomal protein L2 (rplB) {Mycobacterium smegmatis MC2 155}

Sequence Coverage: 76%

Matched peptides shown in **Bold Red**

```
1 MGIRKYKPTT PRRGASVSD FAEITRSTPE KSLVRPLHGK GGRNAHGRT
51 TRHKGGGHR AYRVIDFRH DKDGVNAKVA HIEYDPNRTA NIALLLHYLDG
101 EKRYIAPQG LKQGDVIESG ANADIKPGNN LPLRNIPAGT VIHAVELRPG
151 GGAKLARSAG VSIQLLGKEG TYAALRMP5G EIRRVDVRCR ATVGEVGNAE
201 QSNINWGKAG RMRWKGKRPT VRGVVMNPVD HPHGGGEGKT SGGRHPVSPW
251 GKPEGRTRKP NKPSDKLIVR RRRTGKKR
```

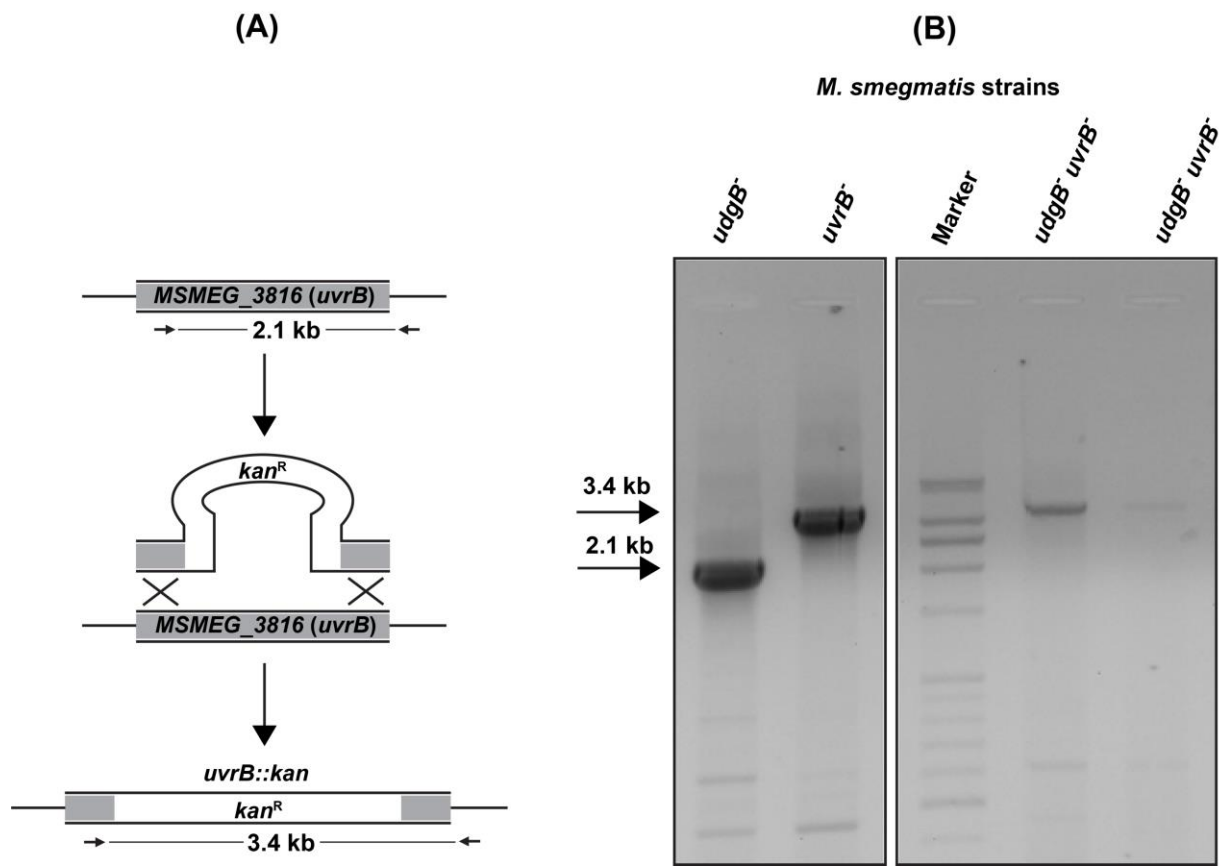
## Search Parameters

Type of search : MS/MS Ion Search  
Enzyme : Trypsin  
Fixed modifications : Carbamidomethyl (C)  
Variable modifications : Oxidation (M)  
Mass values : Monoisotopic  
Protein Mass : Unrestricted  
Peptide Mass Tolerance :  $\pm 0.01$  %  
Fragment Mass Tolerance :  $\pm 0.5$  Da  
Max Missed Cleavages : 0

### **Supplementary References:**

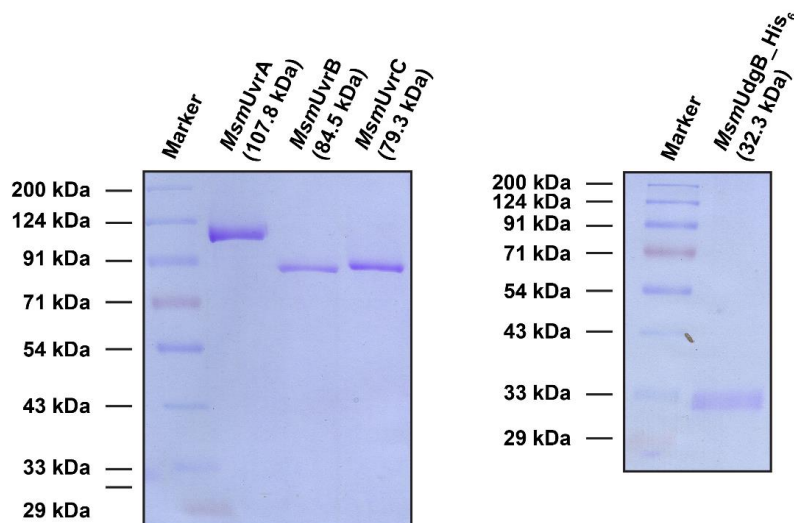
- [1] U. Ray, S. Sharma, I. Kapoor, S. Kumari, V. Gopalakrishnan, S.V. Vartak, N. Kumari, U. Varshney, S.C. Raghavan, G4 DNA present at human telomeric DNA contributes toward reduced sensitivity to gamma-radiation induced oxidative damage, but not bulky adduct formation, *Int J Radiat Biol*, 97 (2021) 1166-1180.
- [2] T. Srinath, S.K. Bharti, U. Varshney, Substrate specificities and functional characterization of a thermo-tolerant uracil DNA glycosylase (UdgB) from *Mycobacterium tuberculosis*, *DNA Repair (Amst)*, 6 (2007) 1517-1528.
- [3] A. China, V. Nagaraja, Purification of RNA polymerase from mycobacteria for optimized promoter-polymerase interactions, *Protein Expr Purif*, 69 (2010) 235-242.
- [4] R.S. Thakur, A. Desingu, S. Basavaraju, S. Subramanya, D.N. Rao, G. Nagaraju, *Mycobacterium tuberculosis* DinG is a structure-specific helicase that unwinds G4 DNA: implications for targeting G4 DNA as a novel therapeutic approach, *J Biol Chem*, 289 (2014) 25112-25136.

Figure S1

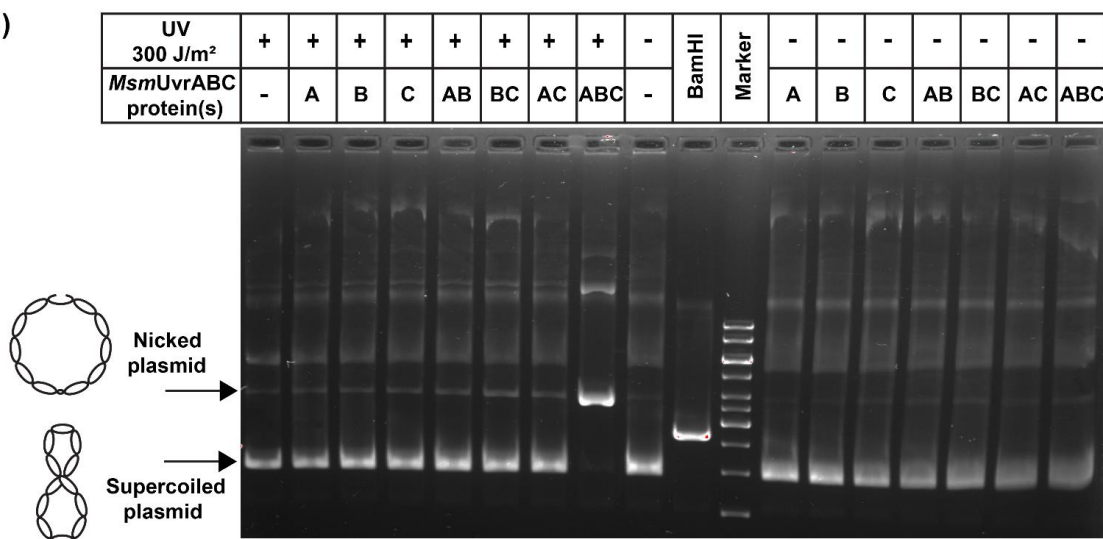


**Fig. S1: Verification of *uvrB* deletion in *M. smegmatis* *udgB*<sup>-</sup> strain.** (A) Schematic showing *MSMEG\_3816 (uvrB)* and *ΔuvrB::kan* loci with the binding sites of screening primers (shown with small horizontal arrows). (B) Representative agarose gel showing the amplification of *uvrB* locus, using the flanking primers, from the control (*udgB*<sup>-</sup>, parent) and knockout strains, where the wild type and *uvrB::kan* alleles resulted in 801 bp and 1594 bp long amplicons, respectively.

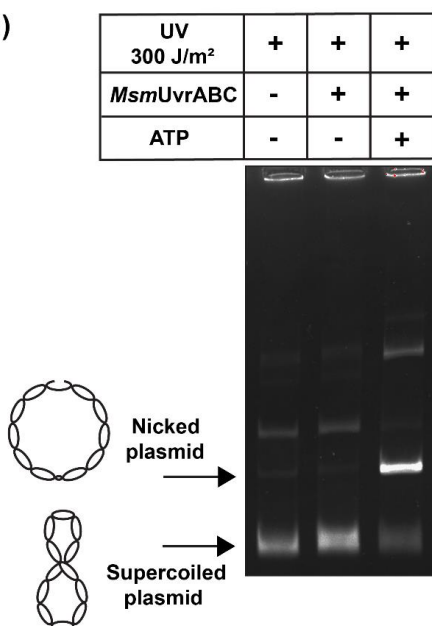
(A)



(B)

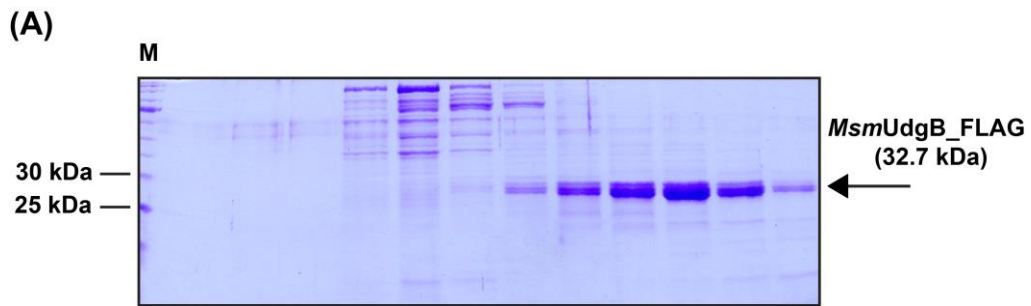


(C)



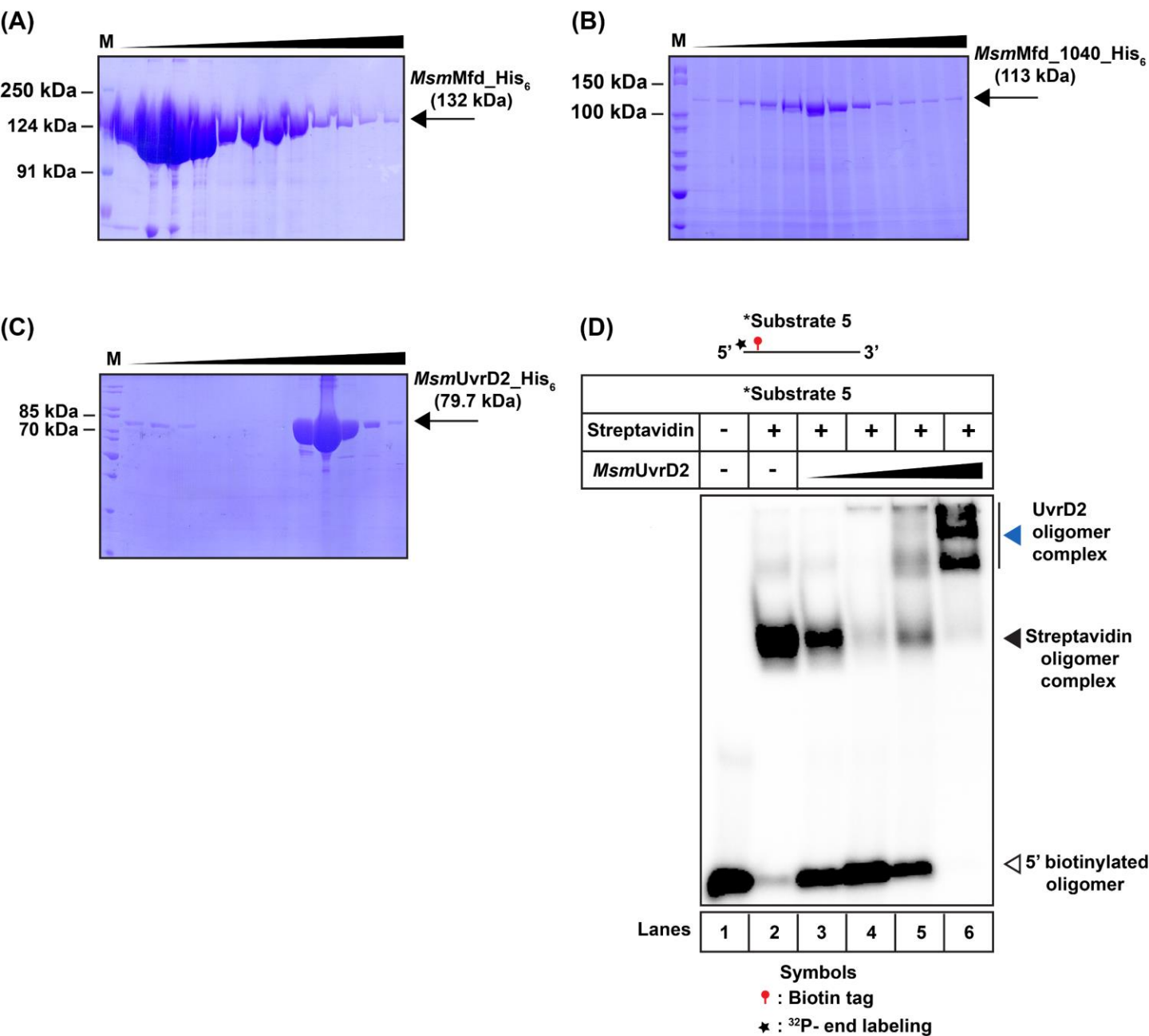
**Fig. S2: Purification and activity analysis of *MsmUvrABC* proteins.** (A) SDS-PAGE showing the purity and migration profile of UvrA, UvrB, UvrC and UdgB. (B) Analysis of the UV-irradiated supercoiled pUC18 (irradiated with 300 J/m<sup>2</sup> UV light) or unirradiated upon their treatments with UvrABC proteins (either one at a time or two at a time or all three proteins together). Resolution of reaction mixtures on 1% agarose gel displays the migration profile of irradiated supercoiled plasmid and its conversion into nicked circular plasmid after treatment with UvrABC complex. (C) Controls for panel (B) examining UvrABC activity in the absence and presence of ATP on the UV-irradiated supercoiled pUC18.

### Figure S3



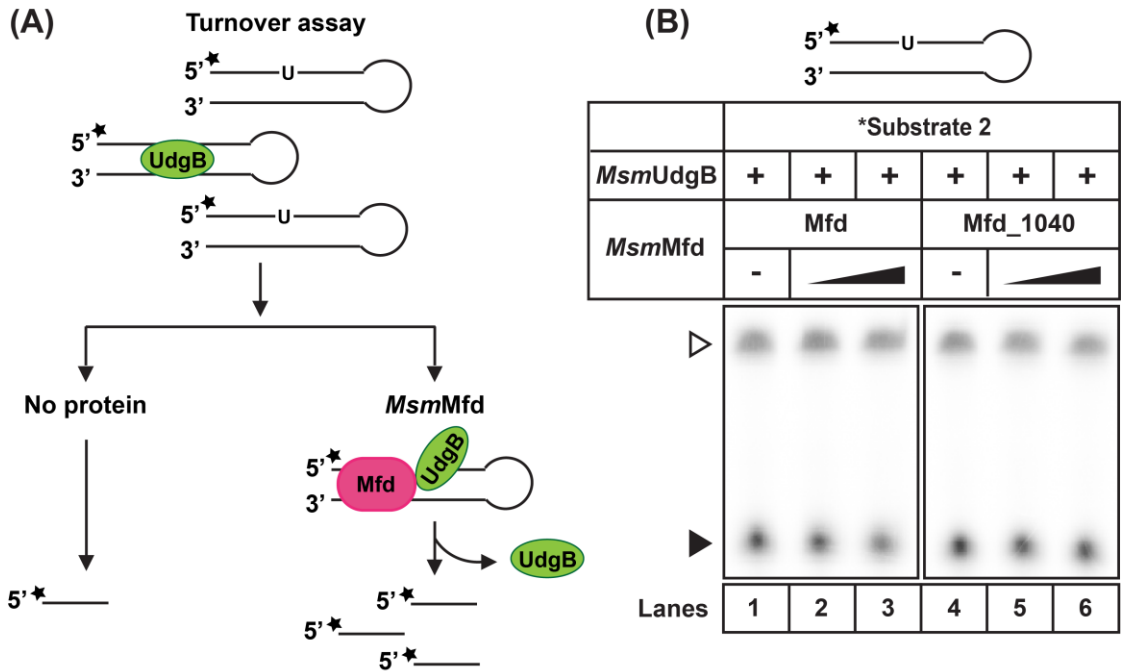
**Fig. S3: Purification profile of *MsmUdgB\_FLAG*.** Representative SDS-PAGE showing the purification (from gel filtration column) and migration profile of UdgB\_FLAG protein containing C-terminal FLAG-tag.

Figure S4



**Fig. S4: Purification of *MsmMfd* and *MsmUvrD2* proteins, and activity analysis of UvrD2.** (A), (B) and (C) SDS-PAGE demonstrating the purification profiles (from Ni-NTA column) and mobility pattern of Mfd, Mfd\_1040 and UvrD2, respectively. (D) Streptavidin displacement assay. Native PAGE depicting the migration of 5' <sup>32</sup>P-labelled and biotinylated substrate 5 (lane 1) and streptavidin complex with the substrate (lane 2). Increasing amounts of the single strand DNA translocase activity of UvrD2 result in streptavidin displacement, migration of substrate at its original position and formation of UvrD2 complex with ssDNA (lanes 3-6). The asterisk denotes 5' <sup>32</sup>P-end labelling of the substrate.

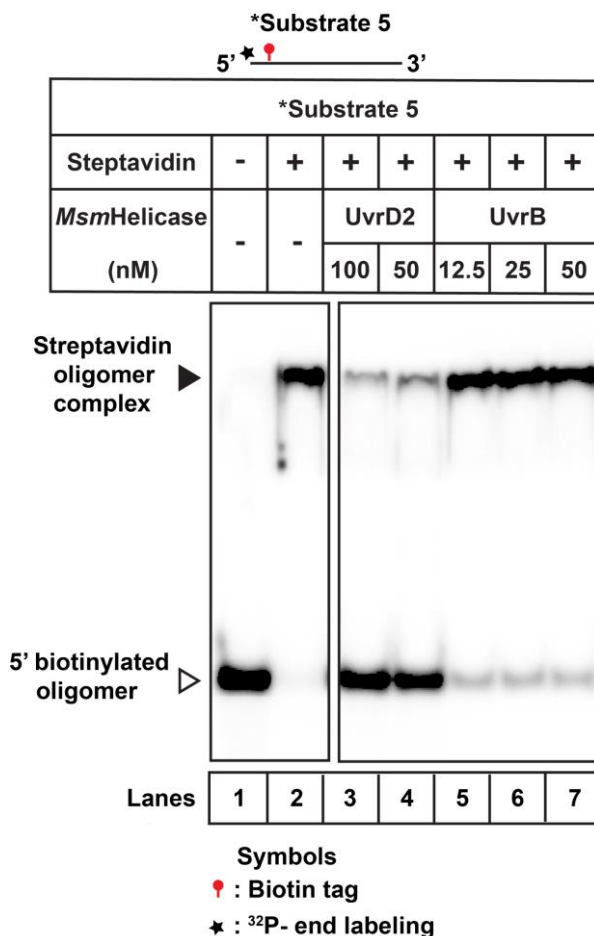
Figure S5



**Fig. S5: Impact of *Msm* translocases on UdgB AP-DNA complex.** (A) Schematic of the turnover assay showing a possible increase in the UdgB reaction product due to the possible translocase action of Mfd (Note: increased product will be seen only if Mfd dislodged UdgB from its complex with AP-DNA). (B) UdgB turnover assays in the presence of either Mfd (full length) or Mfd\_1040 (176 amino acids deleted from C-terminal). <sup>32</sup>P-labelled substrate 2 was treated with the limiting amount of UdgB followed by incubation with the translocase (Mfd or Mfd\_1040). Reaction mixtures from turnover assays were analysed on denaturing PAGE, where the empty black and filled black arrowheads denote <sup>32</sup>P-labelled substrate 2 and UDG reaction product, respectively. The asterisk indicates 5' <sup>32</sup>P-end labelling of the substrate.



**Figure S6**



**Fig. S6: Activity analysis of *Msm*UvrD2 and *Msm*UvrB.** Streptavidin displacement assay. Native PAGE showing the migration of 5' <sup>32</sup>P-labelled and biotinylated substrate 5 (lane 1) and streptavidin complex with the substrate (lane 2). Single strand DNA translocase activity of UvrD2 (lanes 3 and 4) or UvrB (lanes 5-7) results in streptavidin displacement, migration of substrate at its original position. The asterisk denotes 5' <sup>32</sup>P-end labelling of the substrate.