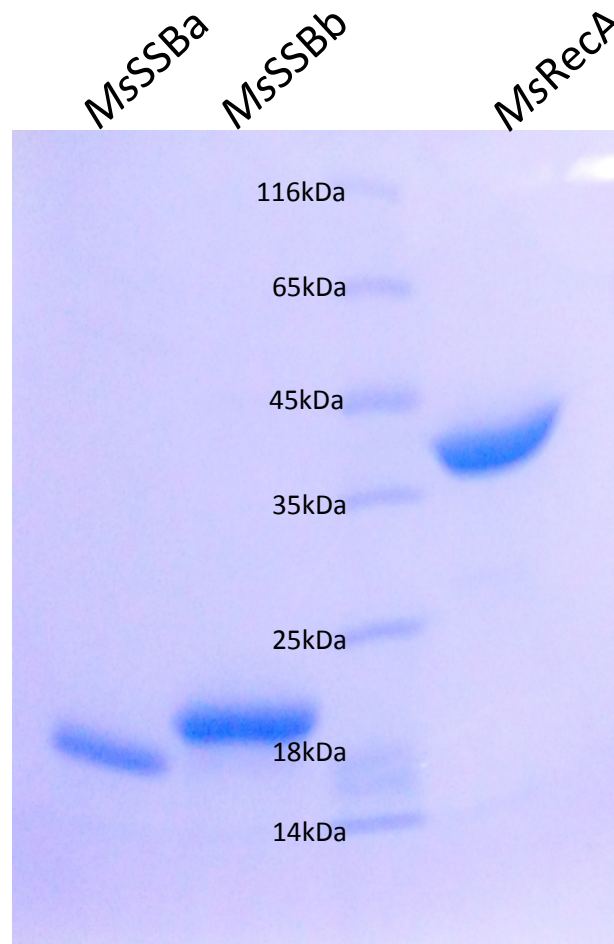
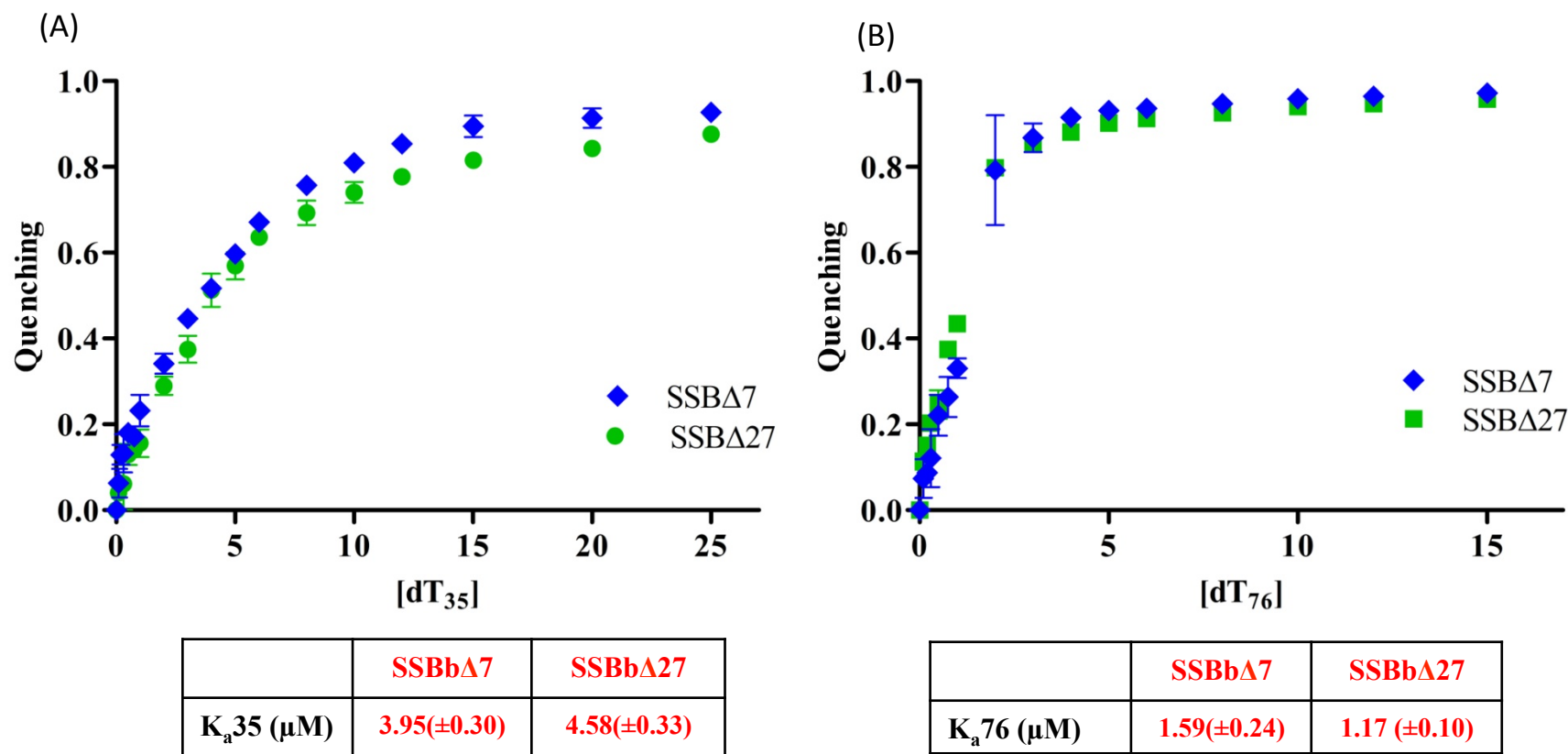


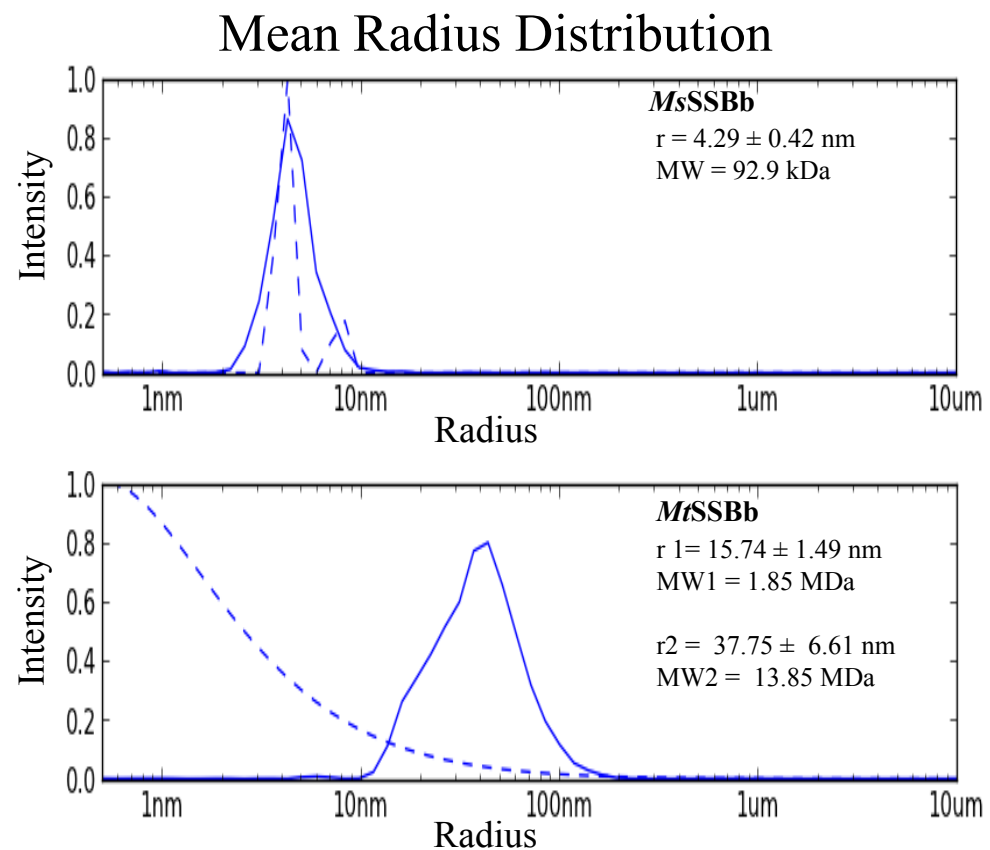
Figure S1



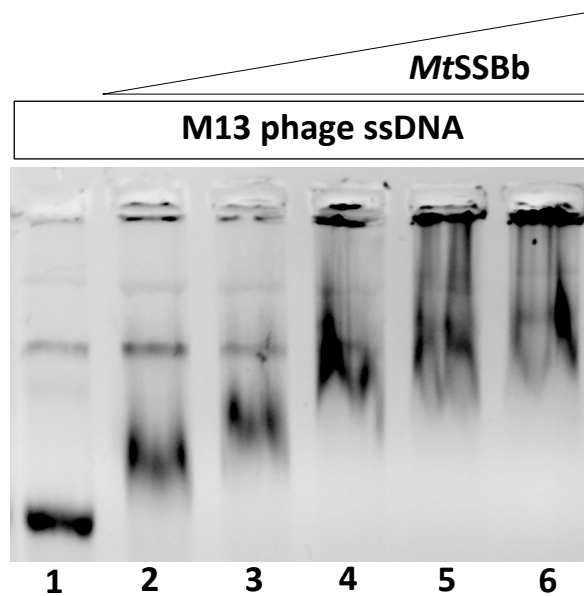
**Figure S1:** SDS PAGE analysis of purified proteins used in the study.



**Figure S2:** Fluorescence studies on truncated SSBb-DNA binding. Titration of 10  $\mu$ M SSB $\Delta$ 7 or SSB $\Delta$ 27 with increasing concentrations of poly-dT<sub>35</sub> (A), or poly-dT<sub>76</sub> (B) in T<sub>20</sub>N<sub>200</sub> [20 mM Tris-HCl pH 8.0, 200 mM NaCl]. [dT<sub>35</sub>] or [dT<sub>76</sub>] represent the concentration of DNA.  $K_a$ 35 and  $K_a$ 76 represent the equilibrium DNA binding constant of poly-dT35 and poly-dT76.



**Figure S3:** Dynamic Light Scattering plot of mycobacterial SSBb. DLS was carried using Spectrosizze 300 (Molecular Dimension). Mean radius distribution plots, showing the probable radius and molecular weight of the following are shown: (A) *MsSSBb*-7mg/ml (B) *MtSSBb*-1mg/ml. Each sample was subjected to 25 readings. Radius distribution plots show a narrow continuous streak for *MsSSBb*, while the map is broad and slightly discontinuous for *MtSSBb*.



**Figure S4:** Analysis of M13 ssDNA binding to *MtSSBb*. M13 ssDNA (250 ng) was incubated with *MtSSBb* (0, 100, 200, 500, 1000, 2000 nmol, lanes 1-6, respectively) in a binding buffer. Reaction mixtures were electrophoresed on 1 % agarose gel (containing EtBr) in 1X TBE. The gel was visualized under UV light.

**Table S1: Primer information for gene cloning**

<i>MsSSBb</i> forward primer	CGTAC <u>ATATGAACATGTT</u> CGAGACACCGTTCACTG
<i>MsSSBb</i> reverse primer	TGAGAAAGCTTCTACGCCGTCAGTGGCAGC
<i>MsSSBb</i> Δ7 reverse primer	TGAGAAAGCTTCTAGCCGGCGAGGTCGGCGACG
<i>MsSSBb</i> Δ27 reverse primer	TGAGAAAGCTTCTAGCGGTCGACGTCATCATCGGGAACG
<i>MsSSBa</i> forward primer	AGCTAC <u>ATATGGCTGGT</u> GACACCACCATCACCGTT
<i>MsSSBb</i> reverse primer	AGCTAAAGCTTCTAGAAGGGCGGTTCGTCGTCAG
<i>MsRecA</i> LIC forward primer	TACTTCCAATCCAATGCAATGGCGCAGCAGGCC
<i>MsRecA</i> LIC reverse primer	TTATCCACTTCCAATGTTATTAGAAAGTCAACCGGG

The restriction enzymes sites added are underlined. Stop codon is colored in red.

**Table S2: Primer information for RT-PCR**

<b>Gene</b>	<b>Size of Amplicon (bp)</b>	<b>Forward Primer</b>	<b>Reverse Primer</b>
<i>ssbB</i> (MSMEG_4701)	102	ATGTTGAGACACCGTTCAC	AATTGCTGGCCACCCGGAAC
<i>ssbA</i> (MSMEG_6896)	124	TGGCTGGTGACACCACCATC	CCAGAGCGGTGAATGGAAAG
<i>sigA</i> (MSMEG_2758)	110	ACCGACGACCTTGAGGTGAC	TTCTTCCTCGTCCTCGACTG
<i>hspX</i> (MSMEG_0424)	124	TTGGGTGCGGGACTTCTTCG	TCCTTGCCGACATCCACACC