Discussion.—From the results (Table I) the following are evident:

- 1. Comparing the activity of compounds 6-10 with compounds 11-15, in the light of their structure at the substituent R", it is clear that the replacement of 'CH₃' group on the cholinic nitrogen atom by 'C₂H₅' group slightly increases the antiacetyl-cholinesterase activity.
- 2. Further, comparing the activity of compounds 1-6 in the light of their structures at R substituent, it is clear that increases in the bulk of R group at 2-position of thiadiazole ring in the title compounds, decreases the anti-acetylcholinesterase activity.
- 3. Change in the position of 'CH₃' group on the phentyl ring substituted on R' (compounds 8-10 and 13-15) is of high significance. As CH₃ group moves from 'o', 'm' to 'p'-position on phenyl ring, the activity increases in title compounds.
- 4. The good anti-acetyl-cholinesterase activity of the title compounds also gives, to some extent, the cholinergic mechanism of excitant action of these compounds on CNS⁴.

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SYNTHESIS OF ELVIROL METHYL ETHER

BECAUSE of the biogenetic interest of the phenolic sesquiterpenoid elvirol¹ (I a), which we suggest, may originate from sesquicarene² (II) by oxidative ring fission (bond a) followed by aromatisation, several syntheses of the phenol and its methyl ether have been reported³. In continuation of our work on phenolic secoscsquiterpenoids⁴, a two step synthesis⁵ of elvirol methyl ether (I b) is described in this communication.

2-Bromo-4-methylanisole (III a) [NMR (60 MHzs $CCl_4 + TMS$): $\delta 2 \cdot 16$ (3H, s, Ar-Me), $3 \cdot 5$ (3H, s, Ar-OM ϵ) and $6 \cdot 53-7 \cdot 33$ (3H, m, Ar-H)] obtained either from p-cresol methyl ether by tromination in glacial acetic acide or from 2-bromo-4-methyl phenology by methylatic n with dimethyl sulphate in alkali, was reacted with lithium dust in ether under nitrogen to furn sh 2-methoxy-5-methylphenyl lithium (III b). Treatment of 6-methylhept-5-en-2-one with the aryllithium (III b) in ether followed by work-up with aqueous hydrochloric acid (1:1), and chromatographic purification of the product (silica gel column-hexane gave the arylheptadiene (IV) in 90% yield [IR ν_{max} (neat): 1620 and 1605 cm⁻¹ (aromatic and C=C); NMR (CCl₄): $\delta 1.70$ (3H, bs,-vinyl-Me), 1.75 (3H,

bs, vinyl-Me), 2·0 (bs, Ar-C-Me), 2·3 (3H, s, Ar-Me), 2·35-3·00 (m, >CH₂), 3·75 (3H, s, Ar-OMe), 4·9-5·4 (m, vinyl-H) and 6·65-7·17 (3H, m, Ar-H)]. Identical diene (IV) (IR, NMR and TLC) was obtained by treatment of methylheptenone with 2-methoxy-5-methylphenylmagnesium bromide (III c).

The regioselective reduction of the styryl double bond in the diene (IV) by lithium in liquid ammonia⁸ (dried over sodium), followed by decomposition with ammonium chloride and chromatographic purification of the product (silica gel column-hexane) gave elvirol methyl ether (I b) in 87% yield [IR (neat): 1605 and 1595 cm⁻¹ (aromatic and C=C); NMR (CCl₄): δ 1·13 (3H, d, J = 7Hz, Ar-CHMe), 1·46 (3H, s, vinyl-Me), 1·60 (3H, s, vinyl-Me), 1·3-2·0 (4H, m, CH₂), 2·2 (3H, s, Ar-Me), 2·8-3·26 (1H, m, Ar-CHMe), 3·7 (3H, s, Ar-OMe), 5·05 (1H, m, vinyl-H) and 6·46-6·93 (3H, m, Ar-H)]. The spectral craracteristics of the specimen agree with those reported for elvirol methyl ether³ (I b). After completion of this work⁵ our attention was drawn to an account of

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the synthesis of elvirol by the Australian workers3 on similar lines.

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CHROMOSOME NUMBER AND DNA, RNA VALUES IN SOME INDIAN BATS (CHIROPTERA)

Our knowledge about the chromosomes of bats has been very meagre till todate despite their wide distribution. A survey of the literature reveals that cytogenetic reports of only a few Indian species of bats are available. The present studies were therefore, undertaken with a view to exploring the chromosomal data on as many representatives of bats as possible. The present findings, which are a part of a larger programme of research work on Indian bats, have been made on nine species. Of these, the chromosomal number of four species, viz., Rhinopoma microphyllum kinneari, Taphozous nudiventris kachensis, Taphozous perforatus perforatus and Hipposideros fulvus pallisidus is being reported for the first time, whereas that for the remaining five species, viz., Cynopterus sphinx sphinx1, Rhinopoma hardwickei hardwickei1-2, Scotophilus heathi heathi3, Megaderma lyra lyra4 and Rousettus leschenaulti5 had been known beforehand hardwickei hardwickei, Iapnozous nuu kachensis

also. The studies also include the estimation of DNA and RNA in mg/gm of known weight of tissues like liver and spleer in order to find out the possible relationship between these biochemical components of the nucleus and the chromosome number in the various species of bats under report.

For chromosomal studies, 0.5% of colchicine per kg body weight was injected intraperitoneally and 21 hours later the specimens were sacrificed and the marrow from the long bones was collected. After hypotonic treatment in sodium citrate (0.9%) for half an hour and fixation in acetic-alcohol (1:3) the usual air-drying technique7 was overnight⁶, followed.

The quantitative estimation of DNA and RNA contents from the tissues was confined to a known weight (100 mg) by employing perchloric acid method⁸⁻⁹. The readings for DNA and RNA were taken from at least five different samples of the same species for every tissue on Bausch and Lomb Spectronic-20. The mean values in mg/gm were thus calculated from the standard graphs prepared by using the standard DNA's and RNA's of calf thymus gland and yeast respectively. The standard deviations and standard errors of the 'mean' were also calculated which refer to variation among the averages obtained from different individuals of the same species.

The studies of chromosomal slides reveal the diploid number of chromosomes varying from 34-54 in these various species (Table I). It is 34 in two, 42 in two. 36 in four and 54 in one species. The studies point cut that the 2N number of chromosomes varies not only in the species belonging to different genera but also in the species of the same genus. Thus the species are rather indistinguishable on the basis of chromosome number alone.

The mean values for DNA, RNA in mg/gm. of the known weight of the various ti-sues obtained through repeated experimentation are also mentioned in Table I and Figs. 1 and 2.

Considering the chromosome number and DNA and RNA values on a collective basis, one finds that the species with the same diploid number of chromosomes possess a variable amount of DNA and RNA in their tissues. This has also been reported in the various species of the genus Bufo and in Amphibia in general10 where the lotal nuclear DNA amount is the most variab, cytogenetic character while the chromosome numbers are relatively constant. Similarly, in the two species, viz., Hipposideros fulvus pallisidus (Microchiroptera) and Cynopterus sphinx sphinx (Megacniroptora) both with a diploid number NA values are higher in of 34 chromc the former th.